This article was downloaded by: On: 23 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



# Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713617200

# An Approach to the Synthesis of Four *Rhodomicrobium* Vannielii Lipid a Analogues

P. Westerduin<sup>a</sup>; T. Beetz<sup>b</sup>; M. J. Dees<sup>a</sup>; C. Erkelens<sup>a</sup>; P. Smid<sup>a</sup>; H. Zuurmond<sup>a</sup>; C. A. A. Van Boeckel<sup>b</sup>; J. H. Van Boom<sup>a</sup>

<sup>a</sup> Gorlaeus Laboratories, RA Leiden, The Netherlands <sup>b</sup> Organon Scientific Development Group, BH Oss, The Netherlands

**To cite this Article** Westerduin, P. , Beetz, T. , Dees, M. J. , Erkelens, C. , Smid, P. , Zuurmond, H. , Van Boeckel, C. A. A. and Van Boom, J. H.(1988) 'An Approach to the Synthesis of Four *Rhodomicrobium* Vannielii Lipid a Analogues', Journal of Carbohydrate Chemistry, 7: 3, 617 - 644

To link to this Article: DOI: 10.1080/07328308808057555 URL: http://dx.doi.org/10.1080/07328308808057555

# PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

## AN APPROACH TO THE SYNTHESIS OF FOUR RHODOMICROBIUM VANNIELII

### LIPID A ANALOGUES

P. Westerduin<sup>1</sup>, T. Beetz<sup>2</sup>, M. J. Dees<sup>1</sup>, C. Erkelens<sup>1</sup>, P. Smid<sup>1</sup>, H. Zuurmond<sup>1</sup>, C. A. A. van Boeckel<sup>2</sup> and J. H. van Boom<sup>1</sup>

 $^1$ Gorlaeus Laboratories, P.O. Box 9502, 2300 RA Leiden, The Netherlands  $^2$ Organon Scientific Development Group, P.O. Box 20, 5340 BH Oss, The Netherlands

Received March 1, 1988 - Final Form June 17, 1988

#### ABSTRACT

The preparation of four *Rh. Vannielii* Lipid A analogues (*i.e.* compounds 22, 23, 30 and 33) is described. Non-neighbouring group supported introduction of the  $\beta$ -glycosidic linkages was performed by coupling the mannopyranosyl bromide 2 and the 2-azido-2-deoxy-glucopyranosyl bromides 10 and 13 with the suitably protected glycosyl acceptors 3, 4 and 5 in the presence of a heterogeneous silver catalyst, to give compounds 6, 7, 14 and 24, respectively. Selective removal of the allyl group and reduction of the azido functions followed by several 0, N-acylation steps afforded, after complete deblocking, the tri- and disaccharide *Rh. Vannielii* Lipid A analogues 22, 23, 30 and 33.

#### INTRODUCTION

Lipopolysaccharide (LPS), an amphiphatic macromolecule in the outer membrane of Gram-negative bacteria, manifests Oantigenic and endotoxic activities and plays an important role in the pathogenesis of Gram-negative bacterial infections.<sup>1-3</sup> A number of studies demonstrated that Lipid A, the lipophilic moiety of LPS, is responsible for most of the biological properties including pyrogenicity, mitogenicity,

617

macrophage activation and lethal toxicity.<sup>4-7</sup> Moreover,detoxified fractions of Lipid A may cause tumor regression<sup>7,8</sup>. The Lipid A's of Gram-negative bacteria, e.g. Salmonella species and Escherichia coli, consist of a  $\beta(1-6)$ -linked Dglucosamine disaccharide substituted by phosphate groups and ester- and amide-bound fatty acids.<sup>7,9</sup> Several analogues of Lipid A have recently been synthesized by several groups.<sup>10-</sup> 18

In 1983, Holst et al.<sup>19</sup> reported the structure of a low toxic Lipid A fragment of *Rhodomicrobium Vannieli*, which lacked covalently linked phosphate groups, but contained instead a  $\beta$ -D-mannopyranosyl residue (see compound 1). In this study we report<sup>20</sup> the synthesis of *Rh. Vannieli* Lipid A analogues 22, 23, 30 and 33, by using the properly protected synthons 1,6-anhydro-2-azido-2-deoxy-D-glucopyranoside and 2,3,4-tri-0-benzyl-D-mannopyranose. The  $\beta$ -glycosidic bonds in the compounds under investigation were introduced in coupling reactions promoted by insoluble silver salts.

## RESULTS AND DISCUSSION

In view of the structural features of the target molecules, we reasoned that application of a heterogeneous silver salt as a promoter for  $\beta$ -glycosylation reactions would be advantageous. The synthesis of the appropriate protected monosaccharides 2-5 was accomplished taking into account substituent effects which determine the stereochemical outcome of a coupling reaction promoted by a heterogeneous silver salt.<sup>21-24</sup> Particularly, the application of a 2azido-2-deoxy-D-glucose derivative is favourable in our strategy: the azido function at C-2 enhances the  $\beta/\alpha$  ratio in

618



coupling reactions promoted by a heterogeneous silver salt<sup>23</sup> and can be selectively reduced to an amino function under mild conditions. In addition, introduction of the 3-hydroxytetradecanoyl amido function can than be performed in a later stage of the synthesis, thus avoiding side reactions during the activation of the anomeric centre in the coupling process.<sup>25,26</sup>

Synthesis of the monosaccharide derivatives 2-5. 6-0-Acetyl-2,3,4-tri-O-benzyl- $\alpha$ -D-mannopyranosyl bromide 2<sup>27</sup> was prepared in three steps from 1,6-anhydro- $\beta$ -D-mannopyranose.<sup>28</sup> Synthesis of the 2-azido-2-deoxy-D-glucopyranosides 3-5 was accomplished according to Paulsen et al.<sup>29,30</sup> starting from 1,6:2,3-di-anhydro- $\beta$ -D-mannopyranose.<sup>31</sup>

Introduction of the  $\beta$ -glycosidic bonds, synthesis of compounds 14 and 24. Glycosylation of 3 with bromide 2 in the presence of silver silicate<sup>32</sup> at -50°C in a mixture of toluene and dichloromethane afforded disaccharide  $6^{33}$  in 80% yield in a  $\beta/\alpha$  ratio of 9/1. Similar results were obtained using 4 as aglycon, giving disaccharide 7. The  $^{13}$ C and  $^{1}$ H NMR spectra of 7 could be completely assigned by two-dimensional NMR techniques (see FIG. 1 and 2A and B). The  $^{13}$ C resonance at 98.9 ppm with J<sub>CH</sub> 154 Hz confirms the presence of a  $\beta$ mannosidic bond.<sup>34</sup>

Acetolysis of 6 and 7 with acetic anhydride/trifluoroacetic acid<sup>35</sup> afforded the corresponding 1,6-diacetates 8 and 11, respectively. Hydrolysis of the anomeric acetate ester with piperidine in tetrahydrofuran<sup>36</sup> yielded the hydroxy compounds 9 and 12, which were converted into their  $\alpha$ -glycosyl bromides 10 and 13 with oxalyl bromide/N,N-dimethylformamide<sup>37</sup>. Condensation of 10 with the glycosyl acceptor 5, as described above, resulted in the exclusive formation of tri-



FIG. 1.  $^{13}C^{-1}H$  Heteronuclear correlated NMR spectrum of compound 7.

saccharide 14 in 70% yield. Glycosylation of benzyl alcohol with bromide 13, under identical conditions, afforded the key  $\beta$ -glycoside 24 in 75% yield.

Preparation of Lipid A derivatives 22 and 23. We now, having key trisaccharide 14 at our disposal, turned our attention to the introduction of the O,N-acyl functions. Zemplén deacetylation of 14 furnished 15 in a quantitative yield. Selective reduction of the azido functions of 15 was effected by







FIG. 2B. Cross-section of the C-5 resonance in 2A revealing a doublet for C-5 and a triplet for  $-\underline{C}H_2Ph$ .



FIG. 3A. <sup>1</sup>H NMR spectrum of the  $-CH_2CO$  moiety of optical pure 3-(R)-hydroxy-tetradecanoic acid methyl ester in the presence of (+)-2, 2, 2-trifluoro-1-(9-anthryl) ethanol.



FIG. 3B. <sup>1</sup>H NMR spectrum of the  $-CH_2CO$  moiety of racemic 3-(R)-hydroxytetradecanoic acid methyl ester in the presence of the same chiral shift reagens as in 3A.

hydrogen sulfide in pyridine/water<sup>38</sup> to afford 16. Infrared spectrometry of the di-amino derivative 16 revealed complete disappearance of the characteristic azide absorption bands, thus confirming reduction of both azido functions. *N*-Acylation of 16 with 3-(R)-hydroxytetradecanoic acid<sup>39</sup> 17 in the presence of *N*-ethylmorpholine, 1-hydroxybenzotriazole, and dicyclohexylcarbodiimide gave glycolipid 20 in 88% yield.

The optical purity of 3-(R)-hydroxytetradecanoic acid 17 was determined by  $^{\rm l}{\rm H}$  NMR spectroscopy of its methyl ester in

the presence of a chiral shift reagent. A small sample of 17 was treated with diazomethane in dichloromethane. The <sup>1</sup>H NMR (300 MHz) spectrum of the  $\alpha$ -methylene group of 3-(R)-hydroxytetradecanoic acid methyl ester together with the chiral shift reagent<sup>40</sup> (+)-2,2,2-trifluoro-1-(9-anthryl)ethanol is illustrated in FIG.3A. The <sup>1</sup>H NMR spectrum of a racemic sample of the corresponding methyl ester is shown in Fig.3B. It can be seen that addition of the shift reagent to the racemate resulted in enantiomeric splitting. Using this technique optical impurities of less than 5% could be detected.

Acylation of 20 with octanoic anhydride in pyridine in the presence of N, N-dimethylaminopyridine afforded glycolipid 21 in a yield of 92%.

Hydrogenolytic cleavage (Pd/C) of the benzyl groups of 20 and 21 in DMF furnished, Lipid A derivatives 22 and 23 in 89% and 57% yields respectively. The <sup>1</sup>H and <sup>13</sup>C NMR spectra of 22 and 23 thus obtained were consistent with the proposed structures.

Preparation of Lipid A derivatives 30 and 33. The molecules to be synthesized (i.e. compounds 30 and 33) contain an ester function at C-3. For this reason it is necessary to replace the acetate esters at C-6 and C-6' by benzyl ethers. Thus, Zemplén deacetylation of key disaccharide 24 afforded 25. Benzylation of the di-hydroxy derivative 25 with NaH and benzyl bromide in THF in the presence of tetrabutylammonium iodide<sup>41</sup> was straightforward and furnished compound 26 in 80% yield. The introduction of the *O*,*N*-acyl functions was performed as follows. Selective removal of the allyl protective group followed by reduction of the azido group would give compound 28. However, isomerization of the allyl ether into a











prop-1-enyl ether, using 1,5-cyclooctadiene-*bis*[methyldiphenylphosphine]-iridium hexafluorophosphate<sup>42</sup>, failed. Fortunately, treatment of 26 with palladium(II)chloride<sup>43</sup> to remove the allyl group yielded the hydroxy derivative 27, although in a rather low yield of 46%. After selective reduction of the azido function, to give 28, the 2-amino and 3-hydroxyl functions were acylated with the activated ester 19, obtained *in situ* from the reaction of 3-(R)-benzyloxytetradecanoic acid 18<sup>44</sup> with 1,1'-bis(6-(trifluoromethyl)benzotriazolyl)oxalate<sup>45</sup> to give fully protected Lipid A derivative 29 in 62% yield. Finally, hydrogenolytic deprotection of 29 afforded Lipid A derivative 30 in 84% yield. The identity of Lipid A derivative 30 was established by <sup>1</sup>H, <sup>13</sup>C NMR spectroscopy and FAB MS.

Selective introduction of 3-(R)-hydroxytetradecanoic acid on the free 2-amino group of 28 to yield 31 was accomplished using 1-hydroxybenzotriazole and dicyclohexylcarbodiimide. Acylation of 31 with tetradecanoic anhydride in the presence of N,N-dimethylaminopyridine gave fully protected Lipid A derivative 32 in 55% yield. The benzyl groups of 32 were removed by catalytic hydrogenation to afford Lipid A derivative 33 in yield of 98%, the identity of which was corroborated by <sup>1</sup>H, <sup>13</sup>C NMR spectroscopy and FAB MS.

Lipid A derivatives 22, 23, 30 and 33 were used to study their biological activities *in vitro* and *in vivo* assays. Preliminary results<sup>46</sup> indicated that compound 23 was active in a lymphocyte transformation test. Compound 23 and 33 exhibited strong immunostimulating (adjuvant) activity in mice as indicated by an increase in serum antibody level against bovine serum albumin.

# FOUR RHODOMICROBIUM VANNIELII LIPID A ANALOGUES

The above mentioned synthesis of Lipid A derivatives 22, 23, 30 and 33 demonstrates that a combined use of 2-azido-2deoxy-D-glucopyranosyl units<sup>47</sup> as synthons together with heterogeneous silver silicate, as a promoter for  $\beta$ -glycosilation reactions, can be successfully applied in the synthesis of Lipid A derivatives.

# EXPERIMENTAL

General Procedures. Tetrahydrofuran, dioxane and pyridine were dried by heating with CaH2 under reflux and then distilled. Ether, dichloromethane, chloroform and toluene were distilled from  $P_2O_5$ . DMF was stirred with  $CaH_2$ at room temperature and distilled under reduced pressure. Pyridine, DMF and tetrahydrofuran were stored over molecular sieves 4A. Toluene and ether were stored over sodium wire and dichloromethane and chloroform over alumina. Schleicher and Schull DC Fertigfolien F1500 LS254 and Merck-Fertigplatten (Kieselgel 60F 254, 5 x 10 cm) were used for TLC analysis. Compounds were detected by charring with 20% sulfuric acid in methanol, or by spraying with 1% potassium permanganate in 5% aqueous potassium carbonate for compounds containing an alkenic component. Optical rotations were recorded at ambient temperature with a Perkin-Elmer 241 polarimeter. Column chromatography was performed on Kieselgel 60, 70-230 Mesh (Merck). Gel filtration was performed on Sephadex LH20 (Pharmacia).  $^{13}$ C NMR spectra were recorded at 50.1 MHz with a Jeol JNM-FX200 spectrometer. <sup>1</sup>H NMR spectra were recorded on a Bruker WM-300 spectrometer equipped with an ASPECT 2000 computer. Chemical shifts are given in ppm ( $\delta$ ) relative to TMS as internal reference, unless otherwise stated.

1,6-Anhydro-2-azido-2-deoxy-3-0-benzy1-4-0-(6-0-acety1-2,3,4-tri-O-benzyl- $\beta$ -D-mannopyranosyl)- $\beta$ -D-glucopyranose (6). To a stirred solution of 1,6-anhydro-2-azido-2-deoxy-3-0 $benzyl-\beta-D-glucopyranose$  (8 mmol, 2.22 g) in toluene/dichloromethane (20 mL, 3/1, v/v) containing silver silicate (8 g) and molecular sieves 4A (4 g), was added dropwise a solution of 6-0-acety1-2,3,4-tri-0-benzy1-a-D-mannopyranosy1 bromide 2 (7 mmol, 3.89 g) in toluene (10 mL) at  $-50^{\circ}$ . Stirring was continued for 3 h, when TLC analysis  $(CH_2Cl_2/$ acetone, 97/3) revealed the formation of one major product.  $CH_2Cl_2$  (150 mL) was added and the mixture stirred 15 min before filtration over celite. The filtrate was concentrated to dryness and the oily residue was applied to a column of Sephadex LH-20 (150 x 3 cm) in 2/1 CH<sub>2</sub>Cl<sub>2</sub>/MeOH. The combined appropriate fractions were concentrated to dryness and the residual oil was purified by silica gel column chromatography (toluene/acetone, 95/5) yielding 71% of 6- $\beta$  and 9% of 6- $\alpha$  as colourless oils:  $[\alpha]^{20} - 32^{\circ}$  (c 1, CHCl<sub>3</sub>);  $R_F = 0.57$  (toluene:acetone, 5:1); <sup>1</sup>H NMR (CDC1<sub>3</sub>)  $\delta$  1.93 (s, 3H, CH<sub>3</sub>CO), 3.27 (s, 1H, H-2), 3.46 (ddd, 1H, H-5', J<sub>4',5'</sub> 9.6 Hz, J<sub>5',6a'</sub> 2.1 Hz, J5',6b' 5.7 Hz), 3.52 (dd, 1H, H-3', J2',3' 3.0 Hz, J31,41 9.4 Hz), 3.74 (c, 1H, H-3), 3.79 (dd, 1H, H-6b, H5.6b 6.0 Hz, J<sub>6a,6b</sub> 7.3 Hz), 3.91 (c, 1H, H-4), 3.92 (t, 1H, H-4',  $J_{3',4'}=J_{4',5'}$  9.5 Hz), 4.04 (d, 1H, H-2',  $J_{1',2'}$  3.0 Hz), 4.16 (dd, 1H, H-6a,  $J_{5,6a} = 1.1$  Hz,  $J_{6a,6b}$  7.3 Hz), 4.23 (dd, 1H, H-6b',  $J_{6a'6b'} = 11.8$  Hz,  $J_{5',6b'} = 5.7$  Hz), 4.47 (dd, 1H, H-6a',  $J_{5',6a'} = 2.1$  Hz,  $J_{6a',6b'} = 11.8$  Hz), 4.49 (d, 1H, CH2Ph), 4.54 (d, 1H, CH2Ph), 4.55 (d, 1H, CH2Ph), 4.66 (d, 1H,  $CH_2Ph$ ), 4.63 (c, 1H, H-5), 4.69 (s, 1H, H-1'), 4.88 (d, 1H, CH<sub>2</sub>Ph), 4.96 (d, 1H, CH<sub>2</sub>Ph), 5.01 (d, 1H, CH<sub>2</sub>Ph), 5.51 (bs, 1H, H-1), 7.04-7.39 (m, 20H, Ph), <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ 20.80 (<u>C</u>H<sub>3</sub>CO), 59.31 (C-2), 63.37 (C-6'), 64.82 (C-6), 71.24, 72.27, 74.17, 75.21 (4 x CH<sub>2</sub>Ph), 72.28 (C-5), 72.95 (C-4), 73.78 (C-2'), 74.04 (C-5'), 74.15 (C-4'), 77.53 (C-3), 81.90 (C-3'), 98.77 (C-1'), 100.48 (C-1), 127.28-128.42 (4x Ph), 137.33, 137.77, 137.89, 138.76 (4x Cquat. Ph), 170.65 (CO).

Anal. Calcd for  $C_{42}H_{45}O_{10}N_3$ : C, 67.0; H, 6.0. Found: C, 66.9; H, 6.0.

1,6-anhydro-2-azido-2-deoxy-3-0-ally1-4-0-(6-0-acety1-2,3,4-tri-O-benzyl-\$-D-mannopyranosyl)-\$-D-glucopyranose (7). Compound 7 was obtained from bromide 2 and aglycon 4 similarly to the preparation of 6: yield 66%;  $[\alpha]^{20}$  -35° (c 1, CHCl<sub>3</sub>); R<sub>F</sub> 0.63 (toluene/acetone, 5/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  2.03  $(3H, CH_3CO)$ , 3.25 (c, 1H, H-2), 3.46 (ddd, 1H, H-5',  $J_{4',5'} =$ 9.6 Hz, J5',6a' = 5.7 Hz, J5'.6b' = 2.1 Hz), 3.55 (dd. 1H, H-3',  $J_{2',3'} = 3.0$  Hz,  $J_{3',4'} = 9.5$  Hz), 3.69 (c, 1H, H-3), 3.78 (dd, 1H, H-6b,  $J_{5,6b} = 6.0$  Hz,  $J_{6a,6b} = 7.3$  Hz), 3.82  $(ddd, 1H, H-4, J_{4,5} = 1.1 Hz), 3.92 (dd, 1H, H-4', J_{3',4'} =$ 9.5 Hz,  $J_{4',5'}$  = 9.6 Hz), 4.04 (dd, 1H, H-2',  $J_{1',2'}$  = 0.3 Hz,  $J_{2',3'} = 3.0$  Hz), 4.10 (dd, 1H, H-6a,  $J_{5,6a} = 1.2$  Hz), 4.23 (dd, 1H, H-6a', J6a',6b' = 11.7 Hz), 4.45 (dd, 1H, H-6b'), 4.64 (c, 1H, H-5), 4.68 (d, 1H, H-1',  $J_{1',2'} = 0.3$  Hz), 5.512 (bs, 1H, H-1), 4.49-5.05 (6H, 3x  $CH_2Oh$ ), 4.12 (c,  $OCH_2$ ally1), 5.21 (c, 2H,=CH<sub>2</sub>), 5.92 (m, 1H, CH=), 7.03-7.40 (c, 15H, Ph), <sup>13</sup>C NMR (CDC1<sub>3</sub>) & 20.79 (CH<sub>3</sub>CO), 59.19 (C-2), 63.31 (C-6'), 64.68 (C-6), 71.13, 117.56, 133.74 (3x C allyl), 72.12 (C-5), 72.80 (C-4), 73.58 (C-2'), 73.93 (C-5'), 74.05 (C-4'), 77.17 (C-3), 81.79 (C-3'), 98.78 (C-1'),  $J_{CH} = 154$ Hz), 100.33 (C-1, J<sub>CH</sub> = 177 Hz), 71.14, 73.96, 75.16 (3x CH2Ph), 127.25-128.31 (3x Ph), 137.82, 138.69 (3x Cquat Ph), 170.59 (CH<sub>3</sub>CO).

Anal. Calcd for  $C_{38}H_{43}O_{10}N_3$ : C, 65.0; H, 6.1. Found: C, 65.1; H, 5.9.

1,6-Di-O-acetyl-3-O-allyl-2-azido-2-deoxy-4-O-(6-Oacetyl-2,3,4-tri-O-benzyl-β-D-mannopyranosyl)- $\alpha/\beta$ -D-glucopyranose (11). To a stirred solution of 7 (2.1 g, 3.0 mmol) in acetic anydride (58 mL) was added dropwise at 0°C trifluoroacetic acid (1.8 mL). After stirring for 3h at 25°C, TLC analysis (CH<sub>2</sub>Cl<sub>2</sub>-acetone, 97:3) showed conversion of the starting material into 11. The solution was coevaporated with toluene (3x 100 mL) and the residue was purified by silica gel column chromatography (eluent CH<sub>2</sub>Cl<sub>2</sub>-acetone, 99:1) to afford 11 (2.4 g, 99%): [ $\alpha$ ]<sub>20</sub> -19° (*c* 1, CHCl<sub>3</sub>) α-acetate; R F 0.31 (CH<sub>2</sub>Cl<sub>2</sub> : acetone, 97:3) α-acetate; <sup>1</sup>H NMR (CDCl<sub>3</sub>) αacetate δ 4.42 (bs, 1H, H-1'), 6.16 (d, 1H, H-1, J<sub>1,2</sub> = 4.0 Hz); <sup>13</sup>C NMR (CDCl<sub>3</sub>) α-acetate δ 20.76, 20.82, 20.91 (3x CO<u>C</u>H<sub>3</sub>), 62.14 (C-6); 63.42 (C-6'), 90.14 (C-1), 101.59 (C-1'), 168.66, 170.38, 170.70 (3x <u>C</u>OCH<sub>3</sub>).

1,6-Di-O-acetyl-4-O-(6-O-acetyl-2,3,4-tri-O-benzyl- $\beta$ -Dmannopyranosyl)-2-azido-2-deoxy-3-O-benzyl- $\alpha/\beta$ -D-glucopyranose (8). Compound 8 was obtained in the same way as described for the synthesis of 11, starting from 6. The analytical data of 8 are comparable with those of 11.

 $6-0-acetyl-4-0-(6-0-acetyl-2,3,4-tri-0-benzyl-\beta-D$ mannopyranosyl)-3-0-allyl-2-azido-2-deoxy- $\alpha$ -D-glucopyranosyl bromide (13). A solution of compound 11 (2.4 g, 2.99 mmol) in dry dioxane (25 mL) was treated for 16h with piperidine (1.42 mL). Dichloromethane (150 mL) was added and the mixture was washed with diluted hydrochloric acid (50 mL), 0.01 N), water (50 mL), aqueous sodium bicarbonate (50 mL, 10%, w/v) and water (50 mL), dried (MgSO<sub>4</sub>) and concentrated to dryness. The

residual oil was purified by column chromatography (silica gel, eluent  $CH_2Cl_2/acetone$ , 9:1) to afford pure 12 (2.1 g, 92%, RF 0.37 (CH<sub>2</sub>Cl<sub>2</sub>/acetone, 9:1). To a vigorous stirred solution of 12 (2.1 g, 2.75 mmol) in chloroform (25 mL) and N,N-dimethylformamide (3.7 mL) was added a solution of oxalyl bromide in chloroform (8 mL, IM) at  $0^{\circ}C$ . The mixture was stirred for 2 h at  $20^{\circ}$ C when TLC analysis (CH<sub>2</sub>Cl<sub>2</sub>/acetone, 95:5) showed the reaction to be complete. The reaction mixture was diluted with ether (150 mL), filtered and washed with cold aqueous sodium bicarbonate (50 mL, 10%, w/v) and cold saturated NaCl solution (50 mL) and dried (MgSO<sub>4</sub>). Evaporation of the solvent afforded crude 13, which was immediately used for further reaction: yield 1.91 g (84%);  $[\alpha]^{20}$  + 39° (c 1, CHCl<sub>3</sub>); R<sub>F</sub> 0.54 (CH<sub>2</sub>Cl<sub>2</sub>/acetone, 95:5); <sup>1</sup>H NMR (CDC13) & 2.03, 2.06 (2x s, 6H, 2x CH3); 3.41 (ddd, 1H, H-5',  $J_{4',5'} = 9.6 Hz$ ,  $J_{5',6a'} = 3.6 Hz$ ), 3.46-3.55 (c, 2H, H-2, H-4), 3.52 (dd, 1H, H-3',  $J_{2',3'} = 3.0$  Hz,  $J_{3',4'} = 9.2$ Hz), 3.77 (d, 1H, H<sub>6</sub>b,  $J_{6a, 6b} = 9.7$  Hz), 3.79 (d, 1H, H-3), 3.87 (d, 1H, H-2'), 3.91 (t, 1H, H-4'), 4.12 (dd, 1H, H-6a,  $J_{5,6a} = 3.0 \text{ Hz}$ ; 4.16 (d, 1H, H-5); 4.33 (d, 2H, H-6a', H-6b'); 4.36 (s, 1H, H-1'); 4.53-4.95 (c, 6H, 3x CH<sub>2</sub>Ph); 5.12-5.21 (c,  $2H_{1}=CH_{2}$ ); 5.92 (o,  $1H_{1}=CH$ ); 6.32 (d,  $1H_{1}$ ,  $H_{1}$ ,  $J_{1,2}$  = 3.8 Hz); 7.02-7.39 (c, 15H, 3x Ph).

 $6-O-Acetyl-4-O-(6-O-acetyl-2,3,4-tri-O-benzyl-\beta-D-mannopyranosyl)-2-azido-2-deoxy-3-O-benzyl-a-D-glucopyranosyl bromide (10). Compound 9 (1.73 g, 2.13 mmol) was treated as described for the preparation of compound 13: yield 1.85 g (98%); R<sub>F</sub> 0.52 (CH<sub>2</sub>Cl/acetone, 95:5); <sup>1</sup>H NMR (CDCl<sub>3</sub>) & 4.41 (bs, 1H, H-1'), 6.34 (d, 1H, H-1, J<sub>1,2</sub> = 4.2 Hz).$ 

 $Methyl-O-(6-O-acetyl-2,3,4-tri-O-benzyl-\beta-D-mannopyrano$  $sy1) - (1-4) - 0 - (6-0 - acetyl - 2 - azido - 2 - deoxy - 3 - 0 - benzyl - \beta - D - benzyl - \beta - benzyl - \beta - benzyl - \beta - benzyl - benzyl$  $glucopyranosyl) - (1-6) - 2 - azido - 2 - deoxy - 3, 4 - di - 0 - benzyl - \beta - D$ glucopyranoside (14). A mixture of alcohol 5 (1.88 mmol), silver silicate (2 g) and 4A molecular sieves (1 g) in dichloromethane (5 mL) was stirred for 1 h at 20°C. Then the mixture was cooled to  $-20^{\circ}$ C and a solution of glycosyl bromide 10 (2.0 mmol) in dichloromethane (3 mL) was added dropwise. The mixture was stirred for 3 h at  $-20^{\circ}$ C, when TLC indicated the formation of 14. Dichloromethane (100 mL) was added to the mixture, solids were removed by filtration and the clear filtrate was concentrated to dryness. The residual oil was applied to a column of Sephadex LH-20 (120 x 3  $cm^2$ ) eluted with  $CH_2Cl_2/MeOH$ , 2:1. The appropriate fractions were collected, concentrated and the residue was chromatographed on Kieselgel 60 (CH<sub>2</sub>Cl<sub>2</sub>/acetone, 97:3 - 95:5) to give 14 as a colourless oil: yield 1.35 g (60%); RF 0.46 (CH<sub>2</sub>Cl<sub>2</sub>/acetone, 95:5); [a]<sup>20</sup> -25° (c 1, CHCl<sub>3</sub>); <sup>1</sup>H NMR (360 MHz) (CHCl<sub>3</sub>) & 2.03, 2.05 (2x s, 6H, 2x  $CH_3$ ); 3.39-3.58 (c, 8H, H-2, H-3, H-4, H-2', H-3', H-5', H-3'', H-5''), 3.61 (ddd, 1H, H-5, J4,5 = 9.5 Hz, J5,6b = 2.1 Hz), 3.65 (s, 3H, OCH3), 3.71 (dd, 1H, H-6a,  $J_{5,6a} = 5.7$  Hz,  $J_{6a,6b} = 11.4$  Hz), 3.86 (dd, 1H, H-4',  $J_{3',4'} = 10$  Hz,  $J_{4',5'} = 8.1$  Hz), 3.92 (t, 1H, H-4'',  $J_{3''}, 4'' = J_{4''}, 5'' = 9.5 Hz$ , 3.93 (bd, 1H, H-2'',  $J_{2''}, 3''$ = 2.8 Hz), 4.18-4.36 (c, 5H, H-6b, H-6a', H-6b', H-6a'', H-6b''), 4.27 (d, 1H, H-1,  $J_{1,2} = 8.0$  Hz), 4.39 (d, 1H, H-1',  $J_{1,2} = 7.9$  Hz), 4.51 (bs, 1H, H-1''), 4.59-5.14 (c, 12H, 6x CH<sub>2</sub>Ph), 7.01-7.42 (c, 30H, Ph); <sup>13</sup>C NMR (CDCl<sub>3</sub>) & 20.47, 20.56 (2x CH<sub>3</sub>), 56.91 (OCH<sub>3</sub>), 62.17 (C-6'), 63.10 (C-6''), 65.68 (C-2'), 66.02 (C-2), 68.50 (C-6), 72.62 (C-5'), 73.50 (C-5''), 73.73 (C-4''), 74.12 (C-5), 74.49 (C-2''), 76.82 (C-4'), 77.61 (C-4), 80.12 (C-3'), 82.08 (C-3''), 82.81 (C-3), 100.68 (C-1), 102.17 (C-1'), 102.67 (C-1''), 71.51, 73.96, 74.24, 74.84, 75.22  $(6x CH_2Ph)$ ; 126.61-128.19 (6x Ph), 137.47-138.32 (6x Cquat Ph), 170.35  $(2x COCH_3)$ .

Anal. Calcd for  $C_{65}H_{72}O_{16}N_6$ : C, 65.4; H, 6.0. Found: C, 65.2; H, 6.0.

Methyl-O-(2,3,4-tri-O-benzyl- $\beta$ -D-mannopyranosyl-(1-4)-O-(2-azido-2-deoxy-3-O-benzyl- $\beta$ -D-glucopyranosyl)-(1-6)-2azido-2-deoxy-3,4-di-O-benzyl- $\beta$ -D-glucopyranoside (15). A catalytic amount of potassium *tert*-butoxide was added to a solution of trisaccharide 14 (0.97 g, 0.81 mmol) in a mixture of methanol (2.5 mL) and dioxane (3.5 mL). After 20 min Dowex 50 H<sup>+</sup>) resin (1 g) was added. The resin was filtered off, washed and the filtrate was concentrated to give 15: yield 98%; R<sub>F</sub> 0.37 (CH<sub>2</sub>Cl<sub>2</sub>/ acetone, 9:1); [ $\alpha$ ]<sup>20</sup> -9° (c 1, CHCl<sub>3</sub>).

Methyl-O-(2,3,4-tri-O-benzyl- $\beta$ -D-mannopyranosyl)-(1-4)-O-(3-O-benzyl-2-deoxy-2-[(R)-3-hydroxytetradecanoylamino]- $\beta$ -D-glucopyrano-syl)-(1-6)-3,4-di-O-benzyl-2-deoxy-2-[(R)-3hydroxytetradecanoyl-amino]- $\beta$ -D-glucopyranoside (20). Compound 15 (0.65 g, 0.59 mmol) was dissolved in pyridine/water (20 ml, 4:1, v/v) and a gentle stream of H<sub>2</sub>S was bubbled through the mixture for 20 h. TLC analysis indicated conversion of the starting material into 16. The reaction mixture was concentrated and the residue was purified by silica gel column chromatography (eluent CH<sub>2</sub>Cl<sub>2</sub>/MeOH/Et<sub>3</sub>N, 90:10:0.05-80:20:0.05) to yield the pure diamino-trisaccharide 16 in 92%. R<sub>F</sub> 0.42 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 9:1). To a solution of 16 (0.57 g, 0.54 mmol), 1-hydroxy-benzotriazole (1.56 mmol), (R)-3hydroxytetradecanoic acid 17 (0.38 g, 156 mmol) and triethylamine (5 mmol) in dioxane (8 mL) dicyclohexylcarbodiimide (1.56 mmol) was added and the mixture was stirred at  $20^{\circ}$ C for 2 h. The solvent was evaporated, the residue was taken up in CH<sub>2</sub>Cl<sub>2</sub> (50 mL) and successively washed with aqueous sodium bicarbonate (25 ml, 10%, w/v) and water (25 mL), dried (MgSO<sub>4</sub>) and concentrated to dryness. The residue was applied to a column of silica gel and eluted with CH<sub>2</sub>Cl<sub>2</sub>/ MeOH (93:7 - 9:1) to give glycolipid 20 (0.72 g, 88%): mp 134-135°; [ $\alpha$ ]<sup>20</sup> -26° (c 1, CHCl<sub>3</sub>); <sup>13</sup>C NMR & 14.29 (2x CH<sub>3</sub>); 37.37, 37.62 (2x  $\alpha$ -CH<sub>2</sub> myristoyl); 55.79, 55.81 (C-2, C-2'), 57.13 (OCH<sub>3</sub>), 61.39 (C-6'), 62.45 (C-6''); 68.40 (C-6); 68.98, 69.11 (2x  $\alpha$ -CH9OH) myristoyl), 80.54 (C-3), 83.04, 83.11 (C-3', C-3''), 101.03 (C-1''), 101.44 (C-1'), 102.60 (C-1), 173.84, 173.86 (2x CO).

Methyl-O-(2,3,4-tri-O-benzyl-6-O-octanoyl- $\beta$ -D-mannopyranosy1)-(1-4)-0-(3-0-benzy1-2-deoxy-2-[(R)-3-octanoy1oxytetradecanoylamino]-6-0-octanoyl- $\beta$ -D-glucopyranosyl)-(1-6)-3,4-di-O-benzyl-2-deoxy-2-[(R)-3-octanoyloxytetradecanoylamino]- $\beta$ -D-glucopyranoside (21). To a solution of compound 20 (0.25 g, 0.165 mmol) in a mixture of  $CH_2Cl_2$  and pyridine (4 mL, 7:1, v/v) were added octanoic anhydride (0.27 g, 1 mmol) and a catalytic amount of DMAP. After 16 h at 20°C, TLC analysis  $(CH_2Cl_2/acetone, 9:1)$  indicated complete conversion of starting material 20 ( $R_F$  0.45) into the desired product 21  $(R_F 0.80)$ .  $CH_2Cl_2$  (50 mL) was added and the mixture was washed with aqueous sodium bicarbonate (25 mL, 10%, w/v) and water (25 mL), dried (MgSO4) and concentrated. The residue was purified on a column of silica gel eluted with CH<sub>2</sub>Cl<sub>2</sub>/acetone, 95:5 - 9:1, to give compound 21 as a waxy solid: yield 0.319 (92%); R<sub>F</sub> 0.8 (CH<sub>2</sub>CL<sub>2</sub>/acetone, 9:1); [a]<sup>20</sup> -6° (c 0.5, CHCl<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>) & 14.05, 14.07 (6x CH<sub>3</sub>); 22.54-31.96 (40x -CH<sub>2</sub>-, fatty acids), 33.91-34.63 (4x CH<sub>2</sub>CO), 41.19-41.92 (2x CH<sub>2</sub>CONH), 56.12, 56.33 (C-2, C-2'); 56.48 (OCH<sub>3</sub>); 63.13, 63.18 (C-6', C-6''); 67.49 (C-6); 70.91, 71.12 (2x CH-OCO); 71.91, 73.21, 73.86, 73.97, 74.02, 74.09, 74.12, 74.36, 75.12, 77.09, 78.22, 80.51 (C-2'', C-3, C-3', C-4, C-4', C-4'', C-5, C-5', C-5'', 6x CH<sub>2</sub>Ph); 82.46 (C-3''); 99.86, 100.74, 101.45 (C-1'', C-1', C-1); 126.70-128.21 (6x Ph); 137.36-138.31 (6x Cquat Ph); 169.31, 169.46 (2x CONH); 173.21, 173.36 (4x CO); <sup>1</sup>H NMR (CDCl<sub>3</sub>) & 0.81-0.91 (c, 18 H, 6x CH<sub>3</sub>), 1.13-1.65 (c, 80 H, 2x CH<sub>2</sub>)<sub>10</sub>, 4x (CH<sub>2</sub>)<sub>5</sub>; 2.11-2.36 (c, 12 H, 6x CH<sub>2</sub>CO), 5.05 (c, 2 H, 2x CHOCO); 5.96, 6.21 (2x d, 2x NH, J<sub>2,NH</sub> = 9Hz); 7.19-7.42 (c, 30 H, 6x Ph).

Methy1-O- $(\beta$ -D-mannopyranosy1)-(1-4)-O-(2-deoxy-2-[(R)-3 $hydroxytetradecanoylamino] - \beta - D - glucopyranosyl) - (1-6) - 2 - deoxy - brain (1-6) - 2 - deoxy - deoxy - brain (1-6) - 2 - deoxy - brain (1-6) - 2 - deoxy - brain (1-6) - 2 - deoxy - d$  $2-[(R)-3-hydroxytetradecanoylamino]-\beta-D-glucopyranoside$  (22). A solution of compound 20 (0.28 g, 0.19 mmol) in ethanol/N,Ndimethylformamide/ acetic acid (20 mL, 14:5:1) was hydrogenated at 20°C and atmospheric pressure in the presence of 10% palladium/charcoal catalyst (200 mg). After 16 h, the catalyst was removed by filtration and the filtrate was concentrated. Purification of the residue on a column of Sephadex LH-20 (45 x 3 cm) in DMF afforded 89% of Lipid A derivative 22.  $[\alpha]^{20}$  -25° (c l, DMF); R<sub>F</sub> 0.15 (CHCl<sub>3</sub>/acetone/  $MeOH/AcOH/H_2O$ , 50:20:10:10:5); mp 218-221°C (dec.); <sup>13</sup>C NMR (DMF-d7) & 14.28 (2x CH<sub>3</sub>); 23.11, 26.20, 26.25, 32.40, 37.60, 37.85 (2x CH<sub>2</sub>)<sub>10</sub>); 44.31, 44.39 (2x CH<sub>2</sub>CO); 56.14, 56.48 (C2, C-2'); 56.51 (OCH<sub>3</sub>); 68.81, 68.89 (2x CHOH); 61.54, 62.52, 68.35, 68.89, 71.89, 73.46, 75.05, 75.91, 76.24, 76.52, 78.51, 81.31 (C-3-C-6, C-3'-C-6', C-2''-C-6''); 102.83 (C-1); 102.73 (C-1'); 102.01 (C-1''); 172.38; 172.76 (2x CONH).

Anal. Calcd for  $C_{47}H_{90}O_{20}N_2$ : C, 56.3; H, 9.0. Found: C, 56.4; H, 8.8.

Methyl-O-(6-0-octanoyl- $\beta$ -D-mannopyranosyl)-(1-4)-O-(2deoxy-2-[(R)-3-octanoyloxytetradecanoylamino]-6-0-octanoyl- $\beta$ -D-glucopyranosyl)-(1-6)-2-deoxy-2-[(R)-3-octanoyloxytetradecanoylamino]-β-D-glucopyranoside (23). Compound 21 (0.22 g, 0.147 mmol) was dissolved in 2-propanol/N,N-dimethylformamide/acetic acid, 14:2:1 and hydrogenated in the presence of 10% palladium on charcoal (200 mg) for 16 h. The mixture was filtered, the filtrate was concentrated and the residue applied to a column of Sephadex LH-20 (45 x 3 cm) in 1:1CH<sub>2</sub>Cl<sub>2</sub> to give pure Lipid A derivative 23 (0.12 g, 57%).  $[a]^{20}$  -13° (c 1, CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 1:1); R<sub>F</sub> 0.51 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 85.15); mp 190-192° (dec); <sup>13</sup>C NMR (CDC1<sub>3</sub>/CD<sub>3</sub>OD, 1:1) & 14.09 (6x CH<sub>3</sub>), 22.71-32.04 (40x (CH<sub>2</sub>)); 34,01, 34.26, 34.63, 34.70 (4x CH<sub>2</sub>CO); 41.40, 41.83 (2x CH<sub>2</sub>CONH); 56.04, 56.22 (C-2, C-2'); 56.53 (OCH3), 62.86 (C-6'), 63.67 (C-6''), 67.37 (C-4''), 68.63 (C-6), 71.53, 71.64 (2x CHOCO); 70.70, 71.02, 72.14, 72.57, 73.71, 74.66, 74.82, 74.99, 81.03 (C-3, C-4, C-5, C-3', C-4', C-5', C-2'', C-3'', C-5''); 100.95 (C-1''); 101.09 (C-1'), 101.76 (C-1); 171.36, 171.78 (2x CONH); 174.25, 174.30, 174.36, 174.50 (4x CO).

Anal. Calcd for  $C_{79H_{146}O_{24}N_{2}}$ : C, 62.9; H, 9.7. Found: C, 63.0; H, 9.9.

Benzyl 6-O-acetyl-4-O-(6-O-acetyl-2,3,4-tri-O-benzyl- $\beta$ -D-mannopyranosyl)-3-O-allyl-2-azido-2-deoxy- $\beta$ -D-glucopyranose (24). A solution of bromide 13 (2.1 g, 2.54 mmol) in toluene (7.5 mL) was added slowly at -65°C to a solution of benzyl alcohol (3.1 mL) in toluene (30 mL) previously stirred for 2 h with silver silicate (3.45 g) and molecular sieves (1.7 g, 4 A) and the mixture was then vigorously stirred for 2 h at  $0^{\circ}$ C. The reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (150 mL), filtered and concentrated. The residue was purified on a silica gel column (CH<sub>2</sub>Cl<sub>2</sub>/acetone, 99:1 - 95:5) to give 24 (1.13 g, 52%): [a]<sup>20</sup> -18° (c 1, CHCl<sub>3</sub>); R<sub>F</sub> 0.59 (CH<sub>2</sub>Cl<sub>2</sub>/acetone, 95:5); <sup>1</sup>H NMR (CDCl<sub>3</sub>) & 3.27 (dd, 1H, H-3, J<sub>2,3</sub> = 9.1 Hz, J<sub>3,4</sub> = 10 Hz), 3.36 (bd, 1H, H-5, J<sub>4,5</sub> = 8.0 Hz), 3.37 (d, 1H, H-2), 3.43 (dd, 1H, H-5', J<sub>4,5</sub> = 9.6 Hz, J<sub>5,6a</sub> = 2.7 Hz), 3.51 (dd, 1H, H-3', J<sub>2',3'</sub> = 2.8 Hz, J3',4' = 9.3 Hz), 3.67 (dd, 1H, H-4), 3.85 (d, 1H, H-2'), 3.88 (t, 1H, H-4', J = 9.5 Hz, 4.15 (dd, 1H, H-6a, J<sub>6a,6b</sub> = 12 Hz, J<sub>5,6a</sub> = 4.5 Hz), 4.25 (d, 1H, H-1, J<sub>1,2</sub> = 8.0 Hz), 4.27 (d, 1H, H-6b), 4.28 (s, 1H, H-6b'), 4.31 (d, 1H, H-6a), 4.43 (bs, 1H, H-1'); 5.08, 5.19, 5.92 (-CH=CH<sub>2</sub>); 4.54-4.96 (8H, 4x CH<sub>2</sub>Ph); 7.02-7.40 (20H, 4x Ph).

Benzyl 3-O-allyl-2-azido-2-deoxy-6-O-benzyl-4-O-(2,3,4,6-tetra-O-benzyl- $\beta$ -D-mannopyranosyl)- $\beta$ -D-glucopyranose (26). To a solution of 24 (1.13 g, 1.32 mmol) in 1:1 methanol/dioxane (16 mL) potassium tert-butoxide (50 mg) was added and the mixture was stirred for 30 min, neutralized with Dowex 50 (H<sup>+</sup>) resin, filtered and concentrated to give 25 100%, [ $\alpha$ ]<sup>20</sup> -37° (c 1, CHCl<sub>3</sub>). To a stirred solution of crude 25 (2.1 mL) was added benzyl bromide (6.3 mmol) and tetrabutylammonium iodide (0.7 mmol). After 10 min, sodium hydride (6.3 mmol) was added and the mixture was stirred for 3.5 h at 50°C. To the cooled solution was added aq. ammonia (25%, 2 mL), and the separated water layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 x 5 mL). The combined organic layers were neutralized at 0°C with aq. HCl. The organic phase was washed with water (2 x 10 mL), dried with MgSO<sub>4</sub>, and concentrated. The residue was applied to a column of silica gel (eluent  $CH_2Cl_2/acetone$ , 100 - 93:7) to give 26 as a colourless oil in 80% yield:  $[\alpha]^{20}$  -34° (c 1, CHCl<sub>3</sub>);  $R_F$  0.55 ( $CH_2Cl_2/acetone$ , 97:3); <sup>1</sup>H NMR (CDCl<sub>3</sub>) & 4.26 (d, 1H, H-1, J<sub>1,2</sub> 8.1 Hz), 4.48 (d, 1H, H-1', J<sub>1',2'</sub> 2.9 Hz); <sup>13</sup>C NMR (CDCl<sub>3</sub>) & 65.82 (C-2), 68.62 (C-6), 69.53 (C-6'); 74.08, 74.78, 75.07, 75.98, 77.64, 81.06, 82.61 (C-3 - C-5, C-2' - C-5'); 100.42 (C-1), 101.12 (C-1'); 69.47, 116.48, 135.40 (Ally1).

Benzyl 2-azido-2-deoxy-6-0-benzyl-4-0-(2,3,4,6-tetra-0benzyl- $\beta$ -D-mannopyranosyl)- $\beta$ -D-glucopyranose (27). To а solution of compound 26 (0.45 g, 0.47 mmol) in acetone/acetic  $acid/H_2O$  (3 mL, 1:1:1) sodium acetate (180 mg) and palladium (II) chloride (105 mg) were added and the mixture was stirred for 24 h at 20°C. The reaction mixture was filtrated, diluted with  $CH_2Cl_2$  (25 mL), washed with water (10 mL), aqueous sodium bicarbonate (10 mL) and water (10 mL), dried (MgSO<sub>4</sub>) and concentrated. The residual oil was applied to a column of silica gel (eluent  $CH_2Cl_2/acetone$ , 99:1 - 97:3) to afford compound 27 as an oil (0.195 g, 46%):  $[\alpha]^{20} - 25^{\circ}$  (c 1, CHCl<sub>3</sub>);  $R_F$  0.48 (CH<sub>2</sub>Cl<sub>2</sub>/acetone, 97:3); <sup>13</sup>C NMR (CDCl<sub>3</sub>) & 65.55 (C-2); 68.36, 69.15 (C-6, C-6'); 70.96, 72.18, 73.50, 73.67, 74.23, 75.22 (6x CH2Ph); 73.93, 74.11, 74.34, 74.63, 74.75, 81.58, 82.57 (C-3 - C-5, C-2' - C-5'); 100.42 (C-1); 101.76 (C-1'); 126.29 - 138.40 (6x Ph).

Benzyl 2-amino-2-deoxy-6-0-benzyl-4-0-(2,3,4,6-tetra-0benzyl- $\beta$ -D-mannopyranosyl)- $\beta$ -D-glucopyranose (28). A solution of compound 27 (0.195 g, 0.21 mmol) in pyridine/triethylamine/H<sub>2</sub>0 (6 mL, 4:1:1) was treated with hydrogen sulfide for 2.5 h. The green solution was concentrated, coevaporated with toluene (20 mL), diluted with  $CH_2Cl_2$  and washed with diluted sodium hydroxide (10 mL, 0.01 N) and water (10 mL) dried (MgSO<sub>4</sub>) and concentrated to give compound **28** (0.189 g, 100%). Infrared spectroscopy of a neat sample of **28** confirmed complete reduction of the azido function (no N<sub>3</sub> stretch at 2110 cm<sup>-1</sup>).

Benzyl  $4-O-(2,3,4,6-tetra-O-benzyl-\beta-D-mannopyranosyl)-$ 6-0-benzy1-3-0-[(R)-3-benzyloxytetradecanoy1]-2-deoxy-2-[(R)-3-benzyloxytetradecanoylamino]- $\beta$ -D-glucopyranose (29). To a suspension of 1,1'-bis[6-(trifluoromethyl)benzotriazolyl]oxalate (129 mg, 0.28 mmol) and (R)-3-benzyloxytetradecanoic acid (92 mg, 0.28 mmol) in acetonitrile (3 mL) was added 4dimethylaminopyridine (DMAP, 37 mg, 0.3 mmol). After the reaction mixture was stirred for 1 h at  $20^{\circ}$ C, a solution of compound 28 (97 mg, 0.11 mmol) and DMAP (37 mg) in acetonitrile (2 ml) was added and stirring was continued for an additional 24 h. The reaction mixture was diluted with  $CH_2Cl_2$ (25 mL), washed with aqueous sodium bicarbonate (10 mL, 10% w/v) and water (10 mL), dried (MgSO<sub>4</sub>) and concentrated. The crude product was purified by silica gel column chromatography (eluent CH<sub>2</sub>Cl<sub>2</sub>/acetone, 98:2 - 90:10) and Sephadex LH-20 column chromatography (CH<sub>2</sub>Cl<sub>2</sub>-MeOH, 2:1) to afford 29 as a waxy solid: yield 70 mg (42%);  $[\alpha]^{20}$  -26 (c 1, CHCl<sub>3</sub>);  $R_F$ 0.45 (CH<sub>2</sub>Cl<sub>2</sub>/acetone, 97:3); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  4.42 (d, 1H, H-1,  $J_{1,2} = 8.4 \text{ Hz}$ , 4.46 (bs, 1H, H-1'), 5.22 (dd, 1H, H-3,  $J_{2,3} = 10.5$  Hz,  $J_{3,4} = 9.2$  Hz), 6.02 (d, 1H, NH, J = 8.4 Hz);  $1^{3}$ C NMR (CDC1<sub>3</sub>)  $\delta$  100.16 (C-1), 100.62 (C-1'); 171.23, 171.73 (2x CONH).

 $2-\text{Deoxy}-2-[(R)-3-\text{hydroxytetradecanoylamino}]-3-0-[(R)-3-\text{hydroxytetradecanoyl}]-4-0-(\beta-D-\text{mannopyranosyl})-\alpha/\beta-D-gluco-$ 

pyranose (30). A solution of compound 29 (50 mg, 0.033 mmol) in N, N-dimethylformamide (6 mL) was treated with hydrogen in the presence of 10% palladium on charcoal (70 mg) for 48 h. The catalyst was removed by filtration and the solution was concentrated. The residual solid was resolved in t-butanol/ water (5 mL, 4:1) and lyophilized to give Lipid A derivative 30 as a fluffy solid (21 mg, 80%):  $R_F$  ( $\alpha/\beta$ ) 0.22/0.18 (CHCl<sub>3</sub>/ acetone/MeOH/AcOH/H2O, 50:20:10:10:5); FAB-MS for C40H75N014: FAB(+) glycerol:  $m/z [M+H]^+$  794, FAB(-)glycerol:  $m/z [M]^-$ . 793; <sup>1</sup>H NMR (CDCl<sub>3</sub>/CD<sub>3</sub>OD, 1:1) & 4.59 (s, 1H, H-1'), 5.13 (d, 1H, H-1 $\alpha$ , J<sub>1,2</sub> = 3.6 Hz), 5.34 (dd, 1H, H-3 $\alpha$ , J<sub>2,3</sub> = 10.9 Hz,  $J_{34} = 9.0 \text{ Hz}$ ,  $\frac{13}{2} \text{ C NMR}$  (CDC1<sub>3</sub>/CD<sub>3</sub>OD, 1:1)  $\delta$  14.20 (2x CH<sub>3</sub>); 22.90, 25.87, 25.96, 29.58, 29.90, 32.15, 36.97, 37.32 (2x-(CH<sub>2</sub>)<sub>10</sub>); 42.60, 43.42 (CH<sub>2</sub>C(0)0, CH<sub>2</sub>CONH); 52.21 (C-2); 71.06, 61.21 (C-6, C-6'); 66.58 (C-4'); 68.48, 68.80 (2x  $\beta$ -C myristoyl); 70.93, 71.28, 71.57, 74.11, 75.25, 76.80 (C-3-C-5, C-2', C-3', C-5'); 91.40 (C-1a), 99.89 (C-1'); 173.36, 173.42 (2x CO).

Anal. Calcd for  $C_{40}H_{75}O_{14}N$ : C, 60.5; H, 9.5. Found: C, 60.7; H, 9.6.

Benzyl 6-O-benzyl-4-O-(2,3,4,6-tetra-O-benzyl- $\beta$ -Dmannopyranosyl)-2-deoxy-2-[(R)-3-hydroxytetradecanoylamino]- $\beta$ -D-glucopyranose (31). Compound 28 (189 mg, 0.21 mmol) was acylated with (R)-3-hydroxy tetradecanoic acid 17 (72 mg, 0.30 mmol) as described in the synthesis of compound 20. The crude product was purified by silica gel column chromatography (eluent CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 97:3 - 95:5) to give compound 31 as a syrup (214 mg, 90%); [ $\alpha$ ]<sup>20</sup> -26 (c 1, CHCl<sub>3</sub>); R<sub>F</sub> 0.20 (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 97:3); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  54.60 (C-2), 100.80 (C-1), 101,21 (C-1').

 $6-0-benzyl-4-0-(2,3,4,5-tetra-0-benzyl-\beta-D-$ Benzyl mannopyranosyl)-2-deoxy-2-[(R)-3-tetradecanoyloxytetradecanoylamino]-3-0-tetradecanoyl- $\beta$ -D-glucopyranose (32). Compound 31 (214 mg, 0.193 mmol) was treated with myristic anhydride (2.63 mg, 0.60 mmol) analogous to the synthesis of compound 21: yield 163 mg (55%);  $[\alpha]^{20} - 22^{\circ}$  (c 1, CHCl<sub>3</sub>);  $R_{\rm F}$ 0.28 (CH<sub>2</sub>Cl<sub>2</sub>/acetone, 97:3); <sup>1</sup>H NMR (CDCl<sub>3</sub>) & 0.88 (C, 9H, 3x CH<sub>3</sub>), 1.13 - 1.56 (c, 64H, (CH<sub>2</sub>)<sub>10</sub>, 2x (CH<sub>2</sub>)<sub>11</sub>), 2.13 - 2.47 (c, 6H, 3x CH<sub>2</sub>CO); 3.30 (c, 1H, H-5'), 3.35 (dd, 1H, H-3',  $J_{2',3'} = 3.4$  Hz,  $J_{3',4'} = 9.5$  Hz), 3.56 (c, 1H, H-5); 3.60, 3.79 (c, 5H, H-6a, H-6b, H-2', H-6a', H-6b'), 3.87 (t, 1H, H-4', J = 9.5 Hz), 3.98 (t, 1H, H-4, J = 9.2 Hz), 4.12 (q, 1H, H-2,  $J_{1,2} = J_{2,3} = J_{2,NH} = 9.2 Hz$ ; 4.39 (s, 1H, H-1'), 4.52 (d, 1H, H-1, J<sub>1,2</sub> = 9.2 Hz); 5.06 (q, 1H, CH<sub>2</sub>CHOCO), 5.16 (t, 1H, H-3), 5.84 (c1, 1H, NH).  $1^{3}$ C NMR (CDC1<sub>3</sub>) & 14.08 (CH<sub>3</sub>); 22.66, 24.64, 24.94, 25.23, 29.14, 29.32, 29.41, 29.64, 31.89 (CH<sub>2</sub> fatty acid residues); 33.87, 34.40, 41.64 (3x CH<sub>2</sub>CO); 53.44 (C-2); 68.74, 69.41 (C-6, C-6'); 70.05, 71.39, 73.23, 73.47, 73.79, 74.96 (6x CH<sub>2</sub>Ph); 71.04 (CH<sub>2</sub>CHOCO); 71.69 (C-3); 74.43 (C-4, C-2'); 74.61 (C-4'); 75.01 (C-5); 75.57 (C-5'); 82.20 (C-3'); 100.01 (C-1); 100.68 (C-1'); 169.36 (CONH); 173.48, 173.97 (2x CO).

2-Deoxy-2-[(R)-3-tetradecanoyloxytetradecanoylamino]-4-O-( $\beta$ -D-mannopyranosyl)-3-O-tetradecanoyl- $\alpha/\beta$ -D-glucopyranose (33). Compound 32 (163 mg, 0.11 mmol) was treated with hydrogen as described for the synthesis of compound 30. Lyophylization of the crude product afforded Lipid A derivative as a white fluffy solid. Yield 115 mg (100%). R<sub>F</sub> 0.32 (CHCl<sub>3</sub>/acetone/MeOH/AcOH/H<sub>2</sub>O, 50:20:10:10:5): <sup>13</sup>C NMR (CDCl<sub>3</sub>/CD<sub>3</sub>OD, 2:1) & 14.31 (3x CH<sub>3</sub>); 23.13, 25.35, 25.55, 25.79, 29.67, 29.85, 30.13, 32.34 (CH<sub>2</sub> fatty acid residues); 34.81, 34.96 (2x CH<sub>2</sub>CO); 41.47 (CH<sub>2</sub>CONH), 52.85 (C-2); 61.59 (C-6); 62.40 (C-6'); 67.80 (C-4'); 71.19 (C-5); 71.51 (C-3, C-2', CHOCO); 74.40 (C-3'); 75.95 (C-4); 77.17 (C-5'); 91.72 (C-1); 99.93 (C-1'); 171.29 (CONH); 174.38, 175.46 (2x CO).

Anal. Calcd for  $C_{54}H_{111}O_{14}N$ : C, 65.0; H, 11.1. Found: C, 65.1; H, 11.0.

#### REFERENCES

- C. Galanos, O. Luderitz, E.Th. Rietschel and O. Westphal in International Review of Biochemistry: *Biochemistry of Lipids II*, 14, 239 (1977); T.W. Goodwin, Ed.; University Park Press, Baltimore.
- S.G. Wilkinson in Surface Carbohydrates of the Prokaryotic Cell 97 (1977); I. Sutherland, Ed.; Academic Press, London, New York.
- O. Luderitz, M. Freudenberg, C. Galanos, V. Lehmann, E.Th. Rietschel and D.H. Shaw, Curr. Top. Membr. Transp., 17, 79 (1982).
- 4. O. Westphal and O. Luderitz, Angew. Chem., 66, 407 (1954).
- O. Luderitz, C. Galanos, V. Lehmann, H. Mayer, E. Th. Rietschel and J. Weckesser, Naturwissenschaften, 65, 578 (1978).
- M.R. Rosner, J.-Y. Tang, I. Barzilay and H.G. Khorana J. Biol. Chem., 254, 5906 (1979).
- Bacterial Endotoxin, Chemical, Biological and Clinical Aspects, Ed. J.Y. Homma, S. Kanegasaki, O. Luderitz, T. Shiba and O. Westphal, Verlag Chemie, Weinheim, 1984.
- K. Takayama, E. Ribi and J.L. Cantrell, Cancer Research, 41, 2654 (1981).
- 9. E.Th. Rietschel, H.W. Wollenweber, H. Brade, U. Zahringer, B. Lindler, U. Seydel, H. Bradaczek, G. Barnickel, H. Labischinski and P. Giesbrecht, Handbook of Endotoxin, Vol. 1, 187 (1984); R.A. Proctor, Ed.; Elsevier Scientific, Amsterdam.
- C.A. van Boeckel, J.P.G. Hermans, P. Westerduin, J.J. Oltvoort, G.A. van der Marel and J.H. van Boom, Recl. Trav. Chim. Pays-Bas, 102, 438 (1983).
- D. Charon, R. Chaby, A. Malinvaud, M. Mondange and L. Szabo, Biochemistry, 24, 2736 (1985).

- 12. S. Nakamoto, T. Takahashi, K. Ikeda and K. Achiwa, Chem. Pharm. Bull., 33, 4098 (1985).
- 13. M. Kiso, S. Tanaka, M. Tanahashi, Y. Fujishuma, Y. Ogawa and A. Hasegawa, *Carbohydr. Res.*, 148, 221 (1986).
- T. Takahashi, S. Nakamoto, K. Ikeda and K. Achiwa, Tetrahedron Lett., 27, 1819 (1986).
- 15. I. Macher, Carbohydr. Res., 162, 79 (1987).
- 16. M. Kiso, S. Tanaka, M. Fujita, Y. Fujishima, Y. Ogawa, H. Ishida and A. Hasegawa, Carbohydr. Res., 162, 127 (1987).
- M. Imoto, H. Yoshimara, M. Yamamoto, T. Shimamoto, S. Kusumoto and T. Shiba, Bull. Chem. Soc. Jpn., 60, 2197 (1937).
- 18. K. Briner and A. Vasella, Helv. Chim. Acta, 70, 1341 (1987).
- O. Holst, D. Boravink, J. Werkesser and H. Mayer, Eur. J. Biochem., 137, 325 (1983).
- 20. P. Westerduin, J.H. van Boom, C.A.A. van Boeckel and T. Beetz, Carbohydr. Res., 137, C4 (1985).
- H. Paulsen, Angew. Chem. 94, 184; Angew. Chem. Int. Ed. Engl., 21, 155 (1982).
- 22. C.A.A. van Boeckel, T. Beetz and S.F. van Aelst, Tetrahedron Lett., 25, 4097 (1984).
- C.A.A. van Boeckel and T. Beetz, Recl. Trav. Chim. Pays-Bas, 104, 171 (1985).
- C.A. van Boeckel and T. Beetz, Recl. Trav. Chim. Pays-Bas, 104, 174 (1985).
- S. Kusumoto, H. Yoshimura, M. Imoto, T. Shimamoto and T. Shiba, Tetrahedron Lett., 26, 909 (1985).
- C. Diolez, M. Mondange, S. Robert Sarfato, L. Szabo and P. Szabo, J. Chem. Soc. Perkin Trans., 1, 275 (1984).
- 27. H. Paulsen and O. Lockhoff, Chem. Ber., 114, 3102 (1981).
- 28. M. Georges and B. Fraser-Reid, Carbohydr. Res., 127, 162 (1984).
- 29. H. Paulsen and W. Stenzel, Chem. Ber., 111, 2334 (1978).
- 30. H. Paulsen and W. Stenzel, Chem. Ber., 111, 2348 (1978).
- 31. J. Stanek and M. Cerny, Synthesis, 698 (1972).
- 32. H. Paulsen and O. Lockhoff, Chem. Ber., 114, 3102 (1981).

- 33. H. Paulsen and R. Lebuhn, Liebigs Ann. Chem., 1047 (1983).
- 34. K. Bock, J. Lundt and C. Pedersen, Tetrahedron Lett., 1037 (1973); K. Bock and C. Pedersen, J. Chem. Soc. Perkin Trans., 2, 293 (1974); K. Bock and C. Pedersen, Acta Chem. Scand., Ser. B, B29 258 (1975).
- 35. H. Paulsen and O. Lockhoff, Tetrahedron Lett., 18, 4027 (1978).
- V.V. Borin, S.E. Zurabyan and A. Ya. Khorlin, Izv. Akad. Nauk. SSSR, Ser. Khim. 2806 (1981).
- 37. S. Sabesan and R.U. Lemieux, Can. J. Chem., 62, 644 (1984).
- H. Paulsen and A. Bunsch, Justus Liebigs Ann. Chem. 2204 (1981).
- 39. M. Demary, G. Puzo and J. Asselineau, Nouv. J. Chim., 2, 373 (1978).
- 40. W.H. Pirkle and D.L. Sikkenga, J. Org. Chem., 42, 1370 (1977).
- 41. S. Czernecki, G. Georgoulis and C. Provelenghiou, Tetrahedron Lett., 17, 3535 (1976).
- 42. J.J. Oltvoort, C.A. van Boeckel, J.H. de Koning and J.H. van Boom, Synthesis 308 (1981).
- 43. T. Ogawa and S. Nakabayashi, Carbohydr. Res., 93, Cl (1981).
- 44. M. Inage, H. Chaki, M. Dmoto, T. Shimamoto, S. Kusumoto and T. Shiba, *Tetrahedron Lett.*, 2011 (1983).
- 45. K. Takeda, K. Tsuboyama, K. Yamaguchi and H. Ogura, J. Org. Chem., 50, 273 (1985).
- 46. H. van de Berg, unpublished results.
- 47. H. Paulsen and M. Schuller, Liebigs Ann. Chem., 249 (1987).